


# Separate F-Type Plasmids Have Shaped the Evolution of the *H30* Subclone of *Escherichia coli* Sequence Type 131

Timothy J. Johnson,<sup>a</sup> Jessica L. Danzeisen,<sup>a</sup> Bonnie Youmans,<sup>a</sup> Kyle Case,<sup>a</sup> Katharine Llop,<sup>a</sup> Jeannette Munoz-Aguayo,<sup>a</sup> Cristian Flores-Figueroa,<sup>a</sup> Maliha Aziz,<sup>b,c</sup>  Nicole Stoesser,<sup>d</sup> Evgeni Sokurenko,<sup>e</sup> Lance B. Price,<sup>b,c</sup> James R. Johnson<sup>f,g</sup>

Department of Veterinary and Biomedical Sciences, University of Minnesota, St. Paul, Minnesota, USA<sup>a</sup>; Division of Pathogen Genomics, Translational Genomics Research Institute, Flagstaff, Arizona, USA<sup>b</sup>; Department of Occupational and Environmental Health, George Washington University, Washington, DC, USA<sup>c</sup>; Modernising Medical Microbiology Consortium, Nuffield Department of Medicine, John Radcliffe Hospital, University of Oxford, Oxford, United Kingdom<sup>d</sup>; Department of Microbiology, University of Washington, Seattle, Washington, USA<sup>e</sup>; Minneapolis Veterans Health Care System, Minneapolis, Minnesota, USA<sup>f</sup>; Department of Medicine, University of Minnesota, Minneapolis, Minnesota, USA<sup>g</sup>

**ABSTRACT** The extraintestinal pathogenic *Escherichia coli* (ExPEC) *H30* subclone of sequence type 131 (ST131-*H30*) has emerged abruptly as a dominant lineage of ExPEC responsible for human disease. The ST131-*H30* lineage has been well described phylogenetically, yet its plasmid complement is not fully understood. Here, single-molecule, real-time sequencing was used to generate the complete plasmid sequences of ST131-*H30* isolates and those belonging to other ST131 clades. Comparative analyses revealed separate F-type plasmids that have shaped the evolution of the main fluoroquinolone-resistant ST131-*H30* clades. Specifically, an F1:A2:B20 plasmid is strongly associated with the *H30R/C1* clade, whereas an F2:A1:B— plasmid is associated with the *H30Rx/C2* clade. A series of plasmid gene losses, gains, and rearrangements involving IS26 likely led to the current plasmid complements within each ST131-*H30* sublineage, which contain several overlapping gene clusters with putative functions in virulence and fitness, suggesting plasmid-mediated convergent evolution. Evidence suggests that the *H30Rx/C2*-associated F2:A1:B— plasmid type was present in strains ancestral to the acquisition of fluoroquinolone resistance and prior to the introduction of a multidrug resistance-encoding gene cassette harboring *bla*<sub>CTX-M-15</sub>. *In vitro* experiments indicated a host strain-independent low frequency of plasmid transfer, differential levels of plasmid stability even between closely related ST131-*H30* strains, and possible epistasis for carriage of these plasmids within the *H30R/Rx* lineages.

**IMPORTANCE** A clonal lineage of *Escherichia coli* known as ST131 has emerged as a dominating strain type causing extraintestinal infections in humans. The evolutionary history of ST131 *E. coli* is now well understood. However, the role of plasmids in ST131's evolutionary history is poorly defined. This study utilized real-time, single-molecule sequencing to compare plasmids from various current and historical lineages of ST131. From this work, it was determined that a series of plasmid gains, losses, and recombinational events has led to the currently circulating plasmids of ST131 strains. These plasmids appear to have evolved to acquire similar gene clusters on multiple occasions, suggesting possible plasmid-mediated convergent evolution leading to evolutionary success. These plasmids also appear to be better suited to exist in specific strains of ST131 due to coadaptive mutations. Overall, a series of events has enabled the evolution of ST131 plasmids, possibly contributing to the lineage's success.

**KEYWORDS:** *Escherichia coli*, ST131, genomes, plasmids


Received 4 May 2016 Accepted 10 June 2016 Published 29 June 2016

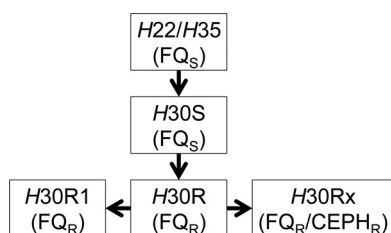
**Citation** Johnson TJ, Danzeisen JL, Youmans B, Case K, Llop K, Munoz-Aguayo J, Flores-Figueroa C, Aziz M, Stoesser N, Sokurenko E, Price LB, Johnson JR. 2016. Separate F-type plasmids have shaped the evolution of the *H30* subclone of *Escherichia coli* sequence type 131. *mSphere* 1(4):e00121-16. doi:10.1128/mSphere.00121-16.

**Editor** Mariana Castanheira, JMI Laboratories

**Copyright** © 2016 Johnson et al. This is an open-access article distributed under the terms of the [Creative Commons Attribution 4.0 International license](https://creativecommons.org/licenses/by/4.0/).

Address correspondence to Timothy J. Johnson, joh04207@umn.edu.

 Separate F-type plasmids have shaped the evolution of the *H30* subclone of *Escherichia coli* sequence type 131 (ST131)



**FIG 1** ST131 subclade emergence and designations, based upon *fimH* allele. FQ<sub>S</sub>, fluoroquinolone sensitive; FQ<sub>R</sub>, fluoroquinolone resistant; CEPH<sub>R</sub>, cephalosporin resistant.

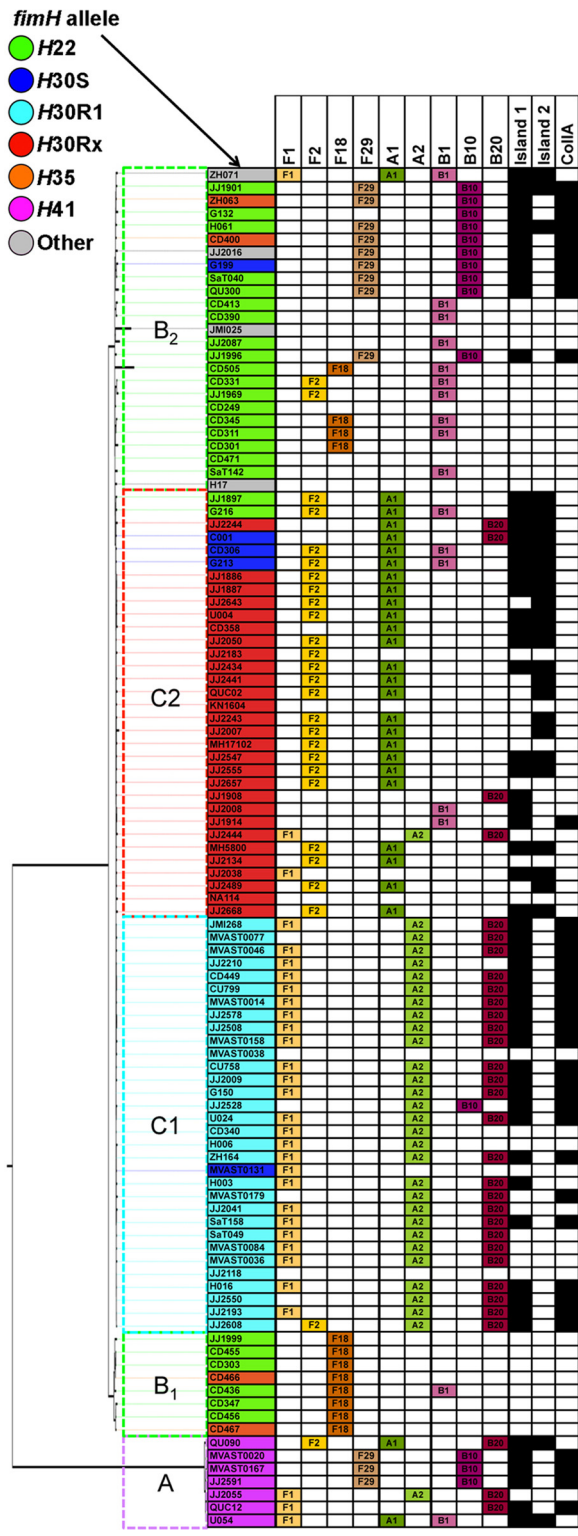
A major global shift has occurred in the distribution of the extraintestinal pathogenic *Escherichia coli* (ExPEC) strains that cause infections of the urinary tract and other extraintestinal sites (1). This shift is best described as an emergence and dominance of *E. coli* sequence type 131 (ST131), a clonal lineage of ExPEC not previously recognized among clinical isolates (2). Within ST131, it is well established that different sublineages have emerged. These sublineages are designated according to their characteristic *fimH* allele (type 1 fimbriae adhesin gene), phylogenetic clade (A, B, C1, and C2, as recently proposed), and resistance profile (3, 4). The most clinically important and expanded of the ST131 sublineages are *H30R1* (where “R” indicates resistance to fluoroquinolones), or clade C1, which has acquired fluoroquinolone resistance; and *H30Rx*, or clade C2, which frequently carries *bla*<sub>CTX-M-15</sub>, conferring extended-spectrum β-lactam resistance (5). *H30R1/C1* and *H30Rx/C2* are sister clades that appear to have evolved from a common ancestor designated *H30R*. The predecessor to *H30R* was *H30S* (where “S” indicates susceptibility to fluoroquinolones), which is thought to have evolved from *H22/B*, a fluoroquinolone- and cephalosporin-susceptible ancestor (Fig. 1) (6, 7). It has been proposed that the *H30Rx/C2* clade emerged in part through acquisition of a plasmid containing *bla*<sub>CTX-M-15</sub> and that this element has subsequently integrated into different chromosomal sites in some strains (5, 6). For simplicity, *fimH* allele designations for the ST131 clonal subsets will be used throughout.

While there is abundant information regarding the clinical aspects and phylogenetics of ST131, the plasmids of ST131 are diverse and not fully defined. F-type plasmids containing multiple plasmid replicons have been identified as dominant within ST131 overall, but their relationships and differential evolution between the various ST131 sublineages are poorly understood (8, 9). The most comprehensive analysis of ST131 plasmids to date (10), which utilized draft genomic sequences, showed that *H30Rx* strains commonly contain an F plasmid with a plasmid multilocus sequence type (pMLST) of F2:A1:B–, based upon the 9 strains examined. It is currently unknown what advantages these plasmids confer besides resistance to antibiotics, although it seems likely that ST131 strains have adapted over time to more efficiently harbor these plasmids (11).

Here, we sought to clarify the within-clade evolution of ST131 in relation to its plasmids by performing comparative plasmid sequencing and analysis of completed plasmids from across the ST131 phylogeny. We also sought to clarify experimentally the *in vitro* transfer potential and stability of different ST131 plasmids.

## RESULTS AND DISCUSSION

**Different F-type plasmids define ST131 sublineages.** A previous collection of 104 diverse ST131 strains described by Price et al. (5) was used first to assess possession of F plasmid allele types by a multilocus analysis approach (Fig. 2). This approach assessed alleles of the FIIA (F), FIA (A), and FIB (B) replicons. Of the 27 *H30Rx* strains, 19 (70%) possessed the F2 allele, 20 (74%) possessed the A1 allele, and 18 (67%) possessed the F2:A1 combination. This was significantly higher ( $P < 0.0001$ ) than the prevalence of F2:A1 plasmids among the 32 *H30R1* strains (0%). In contrast, 25 of the 32 *H30R1* strains (78%) possessed the F1 allele, 30 (94%) possessed the A2 allele, and 26 (81%) possessed



**FIG 2** Distribution of F plasmid allele types, island 1, island 2, and the Colla-containing island among ST131 strains. Strains are ordered by their position in the ST131 phylogenetic tree adapted from reference 5 and colored by *fimH* allele type according to the figure legend. Strain clades (A, B<sub>1</sub>, B<sub>2</sub>, C1, and C2) are designated by colored and labeled dashed boxes. Under island 1, island 2, or Colla, a black-shaded box indicates >99% nucleotide similarity across >90% of the sequence queried.

**TABLE 1** Isolates and plasmids analyzed in this study

Strain	<i>fimH</i> allele	Molecule	Size (bp)	Accession no.	Reference
SaT040	H22	Chromosome	5,061,821	CP014495	This study
		pSaT040	114,223	CP014496	This study
JJ1897	H22	Chromosome	5,174,541	CP013837	This study
		pJJ1897_1	140,502	CP013836	This study
G749	H22	Chromosome	4,897,758	CP014488	This study
		pG749_1	149,732	CP014489	This study
		pG749_2	109,910	CP014490	This study
		pG749_3	62,572	CP014491	This study
CD306	H30S	Chromosome	5,073,822	CP013831	This study
		pCD306	145,221	CP013832	This study
G199	H30S	Chromosome	NA <sup>a</sup>	LSUP00000000	This study
		pG199_1	114,233	LSUP00000000	This study
G150	H30R1	Chromosome	NA	LQHK00000000	This study
		pG150	137,382	LQHK00000000	This study
JJ2434	H30Rx	Chromosome	5,128,614	CP013835	This study
		pJJ2434_1	126,302	CP013833	This study
		pJJ2434_2	62,183	CP013834	This study
ZH063	H35	Chromosome	5,033,359	CP014522	This study
		pZH063_1	114,223	CP014523	This study
		pZH063_2	49,467	CP014524	This study
MVAST0167	H41	Chromosome	4,806,946	CP014492	This study
		pMVAST0167_1	128,305	CP014493	This study
		pMVAST0167_2	78,992	CP014494	This study
MNCRE44	H30R1	pMNCRE44_6	122,966	CP010882	12
JJ1886	H30Rx	pJJ1886-5	110,040	CP006789	13
JJ1887	H30Rx	pJJ1887-4	107,507	CP014321	20
uk_P46212	H30Rx	uk_P46212	143,748	CP013657	6
L46	H30Rx	pEC_L46	144,871	GU371929	14
A	H30Rx	pEK499	117,536	EU935739	15
EC958	H30Rx	pEC958	135,602	HG941719	16
UTI89	ND <sup>b</sup>	pUTI89	114,230	CP000244	17
Uncultured DNA <sup>c</sup>	ND	pRSB225	164,550	JX127248	18
UMN026	ND	p1ESCUM	122,301	CU928148	19

<sup>a</sup>NA, draft sequence where chromosome is in multiple contigs.

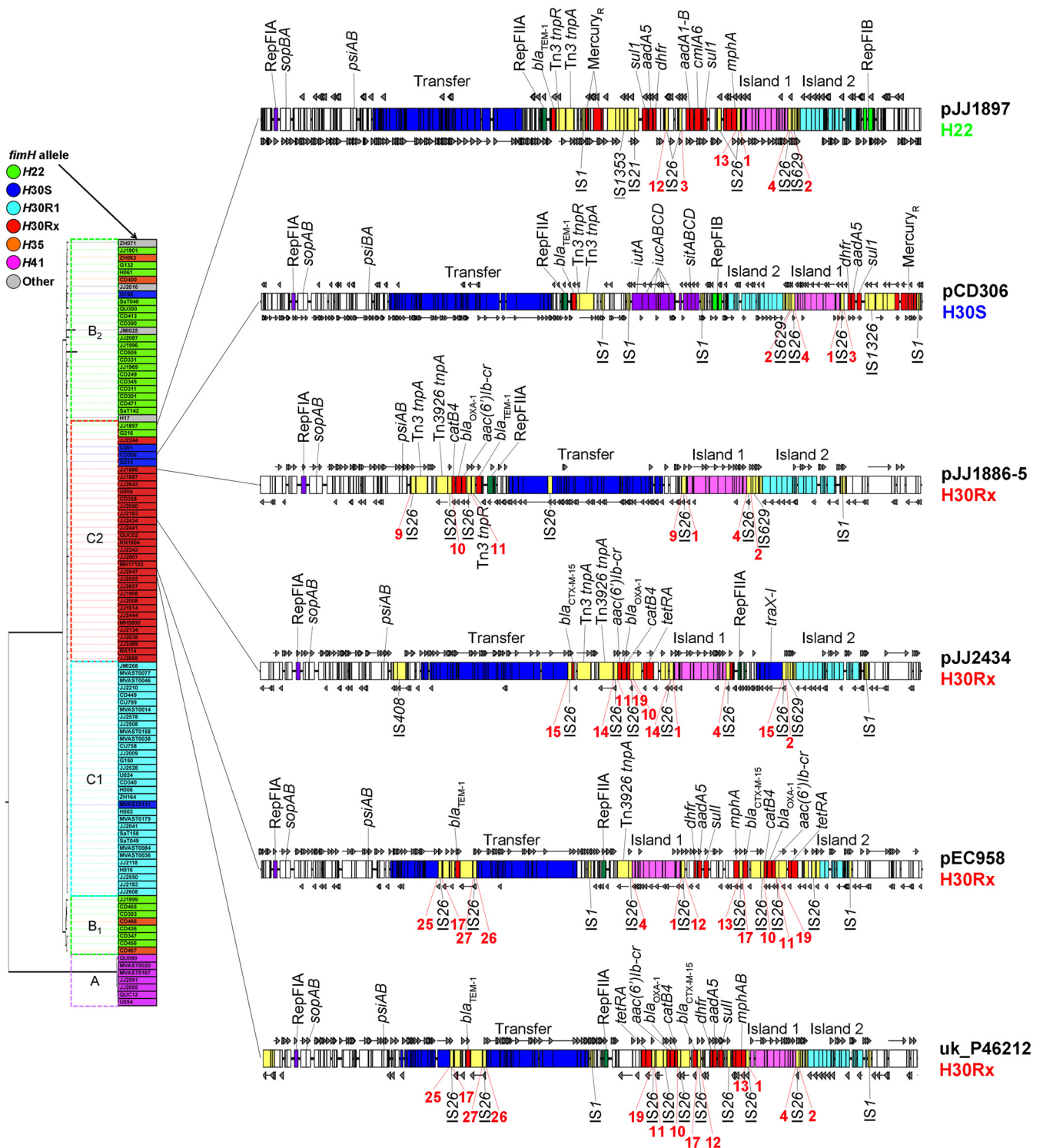
<sup>b</sup>ND, not determined.

<sup>c</sup>The plasmid was not isolated; it was transformed from wastewater sludge plasmid DNA into *E. coli*.

the B20 allele. Thus, the F1:A2:B20 allele combination was fully present in 22 (69%) of the H30R1 strains, versus only 1 (4%) of the H30Rx strains ( $P < 0.0001$ ).

Although F2:A1 (dominating among H30Rx isolates) and F1:A2:B20 (dominating among H30R1 isolates) were not dominant among strains considered ancestral or basal to H30R1 and H30Rx, they nonetheless were identified in several such strains. For example, the F2:A1 allele combination also occurred among H22 and H41 strains, and the F1:A2:B20 allele occurred among H30Rx and H41 strains. However, the more common replicon alleles among ancestral strains were F18, F29, B1, and B10. Specifically, the F29:B10 allele combination was found among H41 and H22, H30S, and H35 strains in the upper portion of the ST131 tree (clade B<sub>2</sub>) and was absent from H30R1 and H30Rx strains (Fig. 2). In contrast, the F18 allele was found among basal H22 and H35 strains in the bottom portion of the ST131 tree (clade B<sub>1</sub>) and was absent from H41, H30R1, and H30Rx strains. Thus, although plasmid allele combinations correlate strongly with strain phylogenetic background, in exceptional instances the same combinations occur in a small number of ancestral isolates. Overall, these data are consistent with acquisition and subsequent loss of F18 and F29:B10 plasmid types during the evolution of ST131, prior to the emergence of ST131-H30.

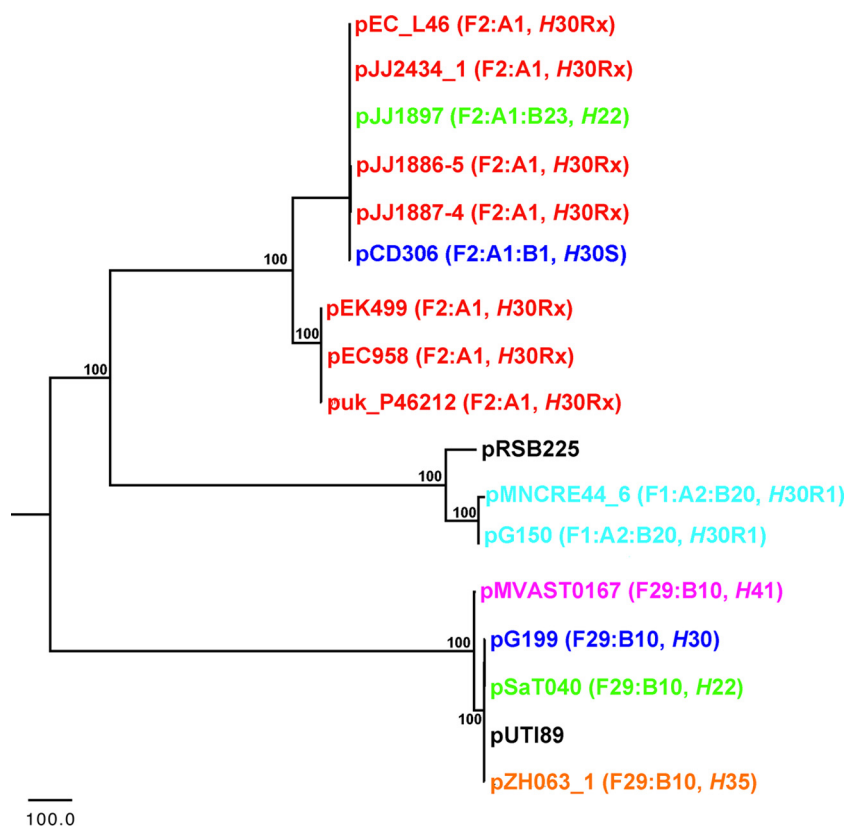
**ST131-H30Rx strains contain an F2:A1 plasmid similar to plasmids from closely related H22 and H30S strains.** The completed sequences of four F2:A1 plasmids from ST131-H30Rx strains, one F2:A1:B23 plasmid from H22, and one F2:A1:B1 plasmid from H30S were examined for their basic structure and orientation of key loci (Table 1; Fig. 3) (6, 12–20). Regardless of host strain clonal background, all six sequenced plasmids contained their expected replicons, stability and maintenance genes



**FIG 3** Linear maps of completed F2:A1 plasmids from H22, H30S, and H30Rx strains. Maps are not to scale. Lines from each map indicate the locations of corresponding isolates in the ST131 phylogenetic tree, with plasmid names colored by *fimH* allele type. Red numbering below each plasmid indicates IS26 tracer sequences, with those of the same number having the same 8-bp flanking sequence.

*psiAB* and *sopAB*, and a conjugative transfer region. They all also possessed two genetic loci that were labeled here as "island 1" and "island 2" (see Fig. S1 in the supplemental material). Island 1 contained nine open reading frames, including a predicted ABC-type transport system associated with iron transport and a putative DNA-binding transcrip-





**FIG 4** Inferred phylogenetic relationships of sequenced ST131 plasmids and closely related database plasmids. Evolutionary history was inferred by the maximum parsimony method, based on 2,414 shared SNP positions among the core regions (32,678 bp) of all plasmids analyzed. Repetitive elements were excluded from the analysis. Isolates are colored by the ST131 sublineage noted in the labels. Numbers at nodes represent bootstrap values based upon 1,000 replicates.

tional regulator. Island 2 contained 12 open reading frames, including a predicted system for carbohydrate transport, another for carbohydrate modification, and a predicted helix-turn-helix (HTH)-type transcriptional regulator.

While these sequenced F2:A1-containing plasmids were generally similar in overall structure, some key differences existed between those from H30Rx strains compared with those from H22 and H30S strains. Specifically (i) only those F2:A1 plasmids from H22 and H30S strains contained an FIB replicon (B23 and B1, respectively), (ii) islands 1 and 2 were inverted in the F2:A1-containing plasmids from H22 and H30S strains, compared to those from H30Rx strains, and (iii) all of the F2:A1 plasmids from H30Rx strains sequenced here, compared to none from the H22 or H30S strains, included a resistance locus containing *aac(6')Ib-cr*, *bla<sub>OXA-1</sub>*, and *catB4*. Notably, numerous copies of IS26 were identified throughout these F2:A1 plasmids, regardless of clonal background. IS26 elements were associated with many rearrangements (discussed below).

Using similarity searches of draft genome sequences, the presence of islands 1 and 2 was sought among all study isolates that contained an F2:A1 plasmid type. Within this subset ( $n = 24$ ), 15 isolates (63%) contained island 1 and 21 (88%) contained island 2. Islands 1 and 2 were also present uniformly among those H22 and H30S isolates (which are considered basal to the H30Rx clade) that exhibited the F2:A1 allelic combination. Whole-plasmid alignments were performed, and phylogenetic inferences were made using conserved single nucleotide polymorphisms (SNPs) in nonrepetitive regions (Fig. 4). This demonstrated that pJJ1897 (H22) and pCD306 (H30S) fell within a monophyletic clade that also included sequenced H30Rx F2:A1 plasmids. Collectively, these results support acquisition of this plasmid type by ST131-H22 prior to the emergence of H30R1 and H30Rx.

**ST131-H30R1 strains contain an F1:A2:B20 plasmid distinct from the plasmids of H30Rx and closely related ST131 strains.** A second F-type plasmid (F1:A2:B20) was identified among H30R1 strains, distinct from the F2:A1 plasmid identified in H30Rx strains (Fig. 5). Key features of the sequenced F1:A2:B20 plasmids included possession of their expected FIA-FIIA-FIB replicons, plasmid stability genes *sopAB* and *psiAB*, and island 1. An additional feature found in all of these plasmids, but lacking in H30Rx plasmids, was a 10-kb region containing the colicin Colla immunity-encoding gene, *cjrABC*, and the enterotoxin-encoding *senB* gene (designated Colla in Fig. 5) (21). Of the 32 H30R1 strains, 23 (72%) contained both island 1 and the Colla-containing region. Another differentiating feature of the F1:A2:B20 plasmids of H30R1, compared to the F2:A1 plasmids of H30Rx, was that they lacked island 2. Based on core plasmid SNPs, the F2:A1:B20 plasmids of H30R1 were clearly distinct from the F2:A1 plasmids of H30Rx, differing by greater than 1,000 core SNPs (Fig. 4).

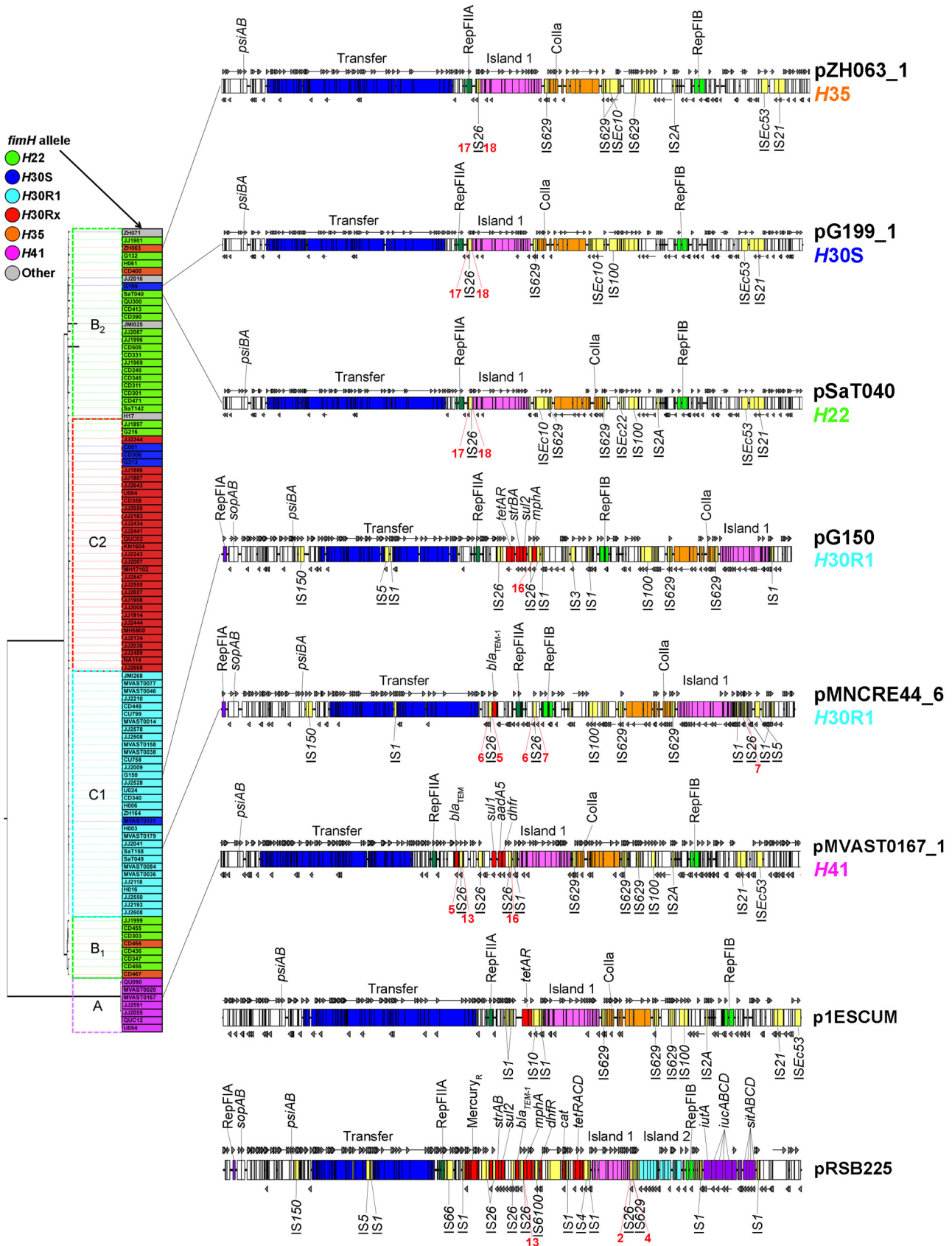
A third plasmid type (F29:B10) was identified among non-H30R1 ST131 strains, including pZH063\_1 (H35), pG199\_1 (H30S), pSaT040 (H22), and pMVAST0167\_1 (H41). These plasmids were highly similar to one another and similar in structure to the F1:A2:B20 plasmids of H30R1 (Fig. 5). The main notable gross differences between the F29:B10 and F1:A2:B20 plasmid types were that the FIA replicon and *sopAB* were absent from the F29:B10 plasmids and the Colla-containing and island 1 regions were in reverse orientation and at different locations on the F29:B10 plasmids. Phylogenetically, however, F29:B10 and F1:A2:B20 plasmids were clearly distinct from one another, thus indicating that at least three plasmid types (F2:A1, F1:A2:B20, and F29:B10) have been introduced into the ST131 clade and harbor some overlapping traits (Fig. 4).

To determine possible origins for the plasmid types, we compared sequenced ST131 plasmids with those in the NCBI reference database (Fig. 4 and 5). Two closely related plasmids were identified harboring similar traits: p1ESCUM from an ExPEC strain (GenBank accession no. [CU928148](https://www.ncbi.nlm.nih.gov/nuclot/CU928148)) and pRSB225 from uncultured wastewater treatment sludge (GenBank accession no. [JX127248](https://www.ncbi.nlm.nih.gov/nuclot/JX127248)). Interestingly, the F1:A2:B20 plasmids from H30R1 strains were phylogenetically most similar to pRSB225, which also contained island 2, the aerobactin siderophore system (22), and the Sit iron transport system (23). In contrast, the other Colla-containing plasmids (pZH063\_1, pG199\_1, pSaT040, and pMVAST0167\_1) were phylogenetically most similar to p1ESCUM. This suggests that a pRSB225-like plasmid may have been introduced initially into ST131, serving as a reservoir for aerobactin, Sit, and islands 1 and 2. However, phylogenetic analyses of regions containing island 1 and Colla (data not shown) from sequenced ST131 plasmids demonstrate multiple discrete clades. This suggests multiple introductions of these mobile regions, not a single introduction, supporting the idea that they were brought in multiple times via different plasmid types.

To further confirm the presence or absence of these plasmid lineages within the broader collection of ST131 strains, whole-plasmid BLAST nucleotide similarity searches were performed, and the results are displayed in Fig. 6. Results support the finding that F29:B10 plasmids are distinct from F1:A2 plasmids (Fig. 6A and B) and that the F2:A1 plasmids typical of H30Rx and basal sublineages are highly conserved and distinct from the F1:A2 and F29:B10 Colla-containing plasmids typical of H30R1 and non-H30R, respectively (Fig. 6C and D).

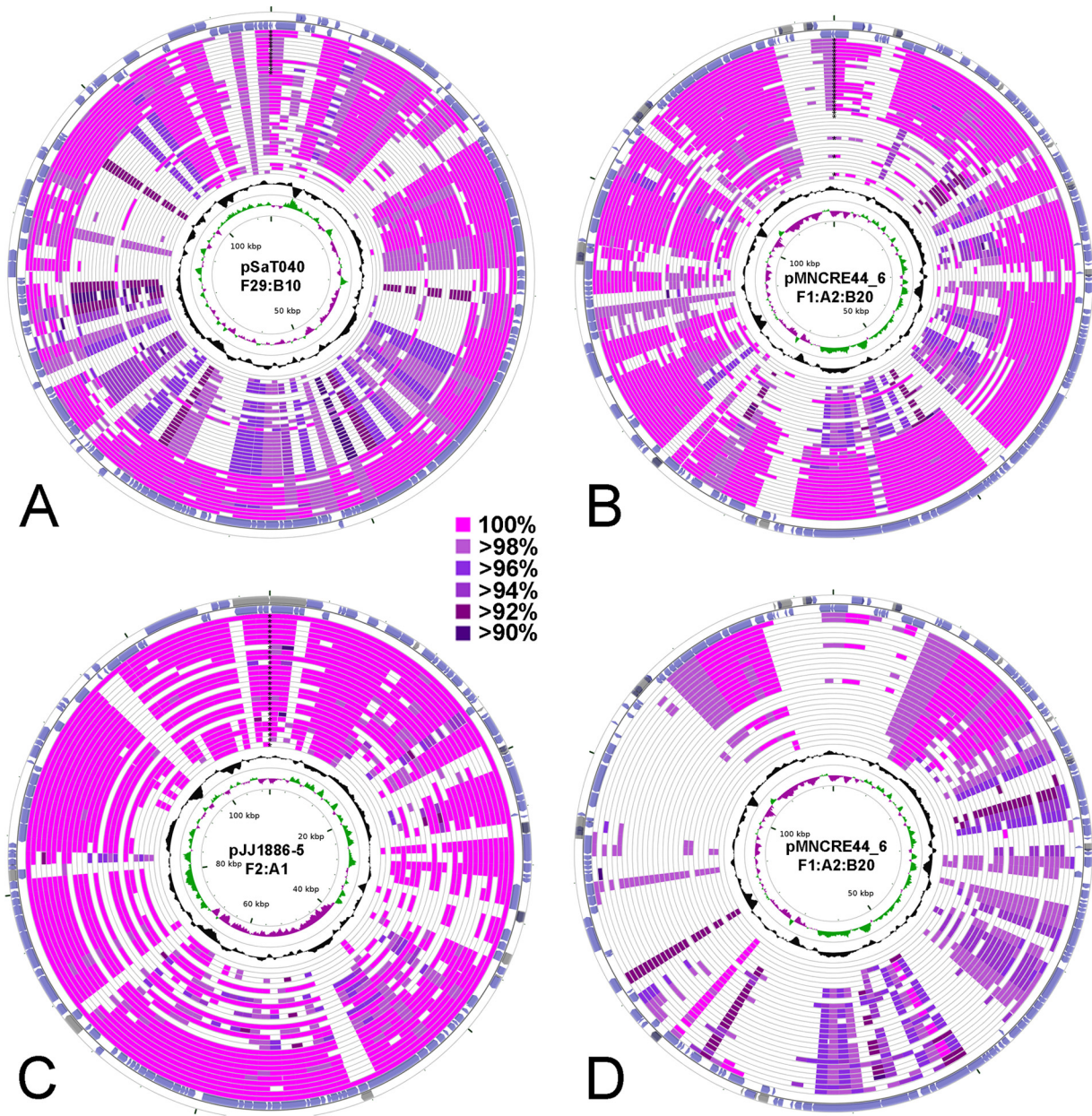
**IS26 actively shapes the microevolution of currently circulating ST131 plasmids.** He et al. recently demonstrated that IS26 plays a major role in plasmid evolution through replicative transposition and cointegrate formation (24). A hallmark of these transposition events is 8-bp target site duplications that are “tracers” of IS26-mediated evolution. Since the ST131 plasmids examined here had high copy numbers of IS26, we performed an analysis of the 8-bp flanking sequences of intact IS26 elements (see Table S1 in the supplemental material).

Similar to the findings of He et al., among more than 50 copies of IS26 examined, we found no left- and right-flanking 8-bp sequences that were identical within an individual IS26 copy (24). Rather, we found that different copies of IS26 within a given plasmid often had identical left- or right-flanking 8-bp sequences, some of which were con-



**FIG 5** Linear maps of completed Colla-containing plasmids from H22, H30S, H35, H41, and H30R1 isolates. Maps are not to scale. Lines from each map indicate the location of the corresponding isolate in the ST131 phylogenetic tree, with plasmid names colored by *fimH* allele type. Red numbering below each plasmid indicates IS26 tracer sequences.





**FIG 6** Circular maps displaying nucleotide conservation of plasmid types across ST131 sublineages. (A) pSaT040 (F29:B10 [accession no. CP014496]) used as reference compared to draft assemblies of Colla-positive isolates. The outer 12 rings are F29:B10-positive isolates, and the inner 27 rings are F1:A2-positive or F29:B10-positive isolates. (B) pMNCRE44\_6 (F1:A2:B20 [accession no. CP010882]) used as a reference compared to draft assemblies of Colla-positive isolates. The outer 21 rings are F1:A2-positive isolates, and the inner 18 rings are F1:A2-positive or F29:B10-positive isolates. (C) pJJ1886-5 (F2:A1 [accession no. CP006789]) used as a reference compared to draft assemblies of all F2:A1-positive isolates. (D) pMNCRE44\_6 (F1:A2:B20 [accession no. CP010882]) used as a reference compared to draft assemblies of all F2:A1-positive isolates. The outer two rings are coding regions corresponding to coordinates in GenBank files. The inner two rings represent G+C content and G+C skew, respectively. Asterisks at the zero coordinates designate plasmids of the same replicon allele combination for each respective reference plasmid.

served between different plasmids. This strongly indicates that rather than intermolecular transposition, the primary driver of IS26-mediated evolution of these plasmids is through (i) *trans* intramolecular transposition resulting in IS26 duplication and segment inversion or (ii) *cis* intramolecular transposition resulting in deletion of plasmid segments. This was evident when comparing closely related plasmids (red numbering on Fig. 3 and 5). There was clear evidence of differentiation between plasmid types (F2:A1, F1:A2:B20, and F29:B10) based upon IS26 flanking 8-bp sequences, with each plasmid allelic type having a distinct subset of flanking sequences that were shared between

plasmids of the same type. For example, identical 8-bp flanking sequences were found for F1:A2:B20 plasmids pZH063\_1 (*H35*), pG199\_1 (*H30S*), and pSaT040 (*H22*) (Fig. 5). Similar 8-bp flanking sequences were also found surrounding island 1 in F2:A1 plasmids (Fig. 3), supporting the common origin of these plasmids.

We also found a logical progression in IS26 8-bp flanking sequences from basal to recent isolates, supporting initial acquisition of IS26 elements followed by extensive intramolecular transposition leading to the current ST131-*H30Rx* strains (Fig. 3). For example, in pCD306 (*H30S*, basal to *H30Rx*), Islands 1 and 2 are downstream of the aerobactin and Sit iron acquisition-related systems, with the FIB replicon positioned between aerobactin-Sit and island 2-island 1. In pJJ1897, aerobactin-Sit is absent and the FIB-island 2-island 1 region is inverted yet retains IS26 8-bp sequences flanking island 1, which were also present in pCD306. This orientation is then conserved in all sequenced *H30Rx* plasmids, except that FIB is absent and instead is replaced by a conserved cluster of hypothetical genes separated from island 2 by an *IS1* copy, supporting the common origin for these plasmids from a basal strain (i.e., *H22*) within the ST131 clade.

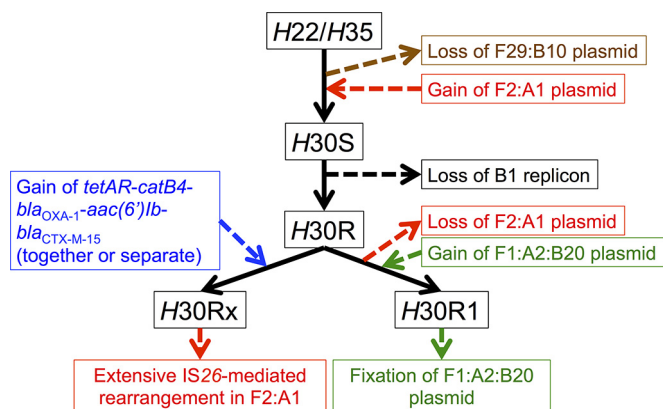
Also, multiple IS26 insertions were found in the F transfer region of sequenced *H30Rx* plasmids, exemplified in simplest form in pJJ1886-5 as an insertion between *tral* and *traH*. This initial insertion likely led to additional IS26-mediated rearrangements in plasmid pJJ2434, for example, where the transfer region was disrupted: *finO-traX-tral* and the RepFIIA replicon were translocated to between islands 1 and 2, a portion of the transfer region was deleted, and the remainder of the transfer region and its adjacent antibiotic resistance island were inverted in the original location. In *H30Rx*-origin plasmids pEC958 and uk\_P46212, a different IS26 insertion in the F transfer region apparently led to the addition of *bla*<sub>TEM-1</sub> within the transfer region. These are only a few examples of the numerous rearrangements and deletions that seemingly have occurred during the evolution of this plasmid within the *H30Rx* clade, particularly within antibiotic resistance-associated islands. Given the compressed time frame of *H30Rx* clade evolution (6), it is evident that IS26-mediated plasmid rearrangements are ongoing in the *H30Rx* clade.

#### **ST131 F-type plasmids have a comparatively low *in vitro* transfer rate.**

Sequence evidence suggests that the plasmids of *H30R1* and *H30Rx* may be fixed within their lineage, rarely transferring between strains. Using representative plasmids and host strains (see Table S2 in the supplemental material), we assessed the transfer frequency of ST131-source F2:A1 and F1:A2:B20 plasmids into several host backgrounds, including laboratory strains DH10B and K-12 MG1655, and (ST131) *H22* to *H30R1* strains lacking plasmids (see Fig. S2 in the supplemental material). Strains containing typical FIIA/FIB, I1, and  $\chi^2$  plasmids were included for comparison as donors representing typical non-ST131 plasmids of different replicon types.

The transfer frequencies of ST131-source plasmids were much lower than those of non-ST131-source plasmids (see Fig. S2 in the supplemental material). In particular, the transfer frequencies of ST131-source plasmids were 2 to 3 log<sub>10</sub> lower than a prototypic F-type ColV plasmid from strain APEC O2 (22, 25). The tested *H30R1* plasmids transferred at higher frequency than the tested *H30Rx* plasmids, possibly due to the innate transfer properties of F2:A1 versus F1:A2:B20 plasmids or due to a helper effect from the additional plasmids harbored by the tested *H30R1* strains tested acting as helpers in conjugation (see Table S2 in the supplemental material). Different host backgrounds did not significantly influence the transfer frequencies of ST131-*H30Rx* plasmids (see Fig. S2). Overall, there appears to be a generalized deficiency in the ability of ST131-*H30Rx* plasmids to transfer spontaneously into recipient hosts that is independent of donor-recipient matching and that likely is due to the observed disruptions of the F transfer region in sequenced *H30Rx* plasmids.

**ST131-*H30Rx* plasmids demonstrate differential *in vitro* stability.** Sequence data also suggested that some strains within the *H30R1* and *H30Rx* sublineages have lost their F-type plasmids, even though such plasmids are thought to be highly stable in the *E. coli* host (8). The *in vitro* stability of the plasmids of strains JJ1886 and JJ1887



**FIG 7** Proposed events leading to currently circulating plasmids in ST131-*H30R1* and *H30Rx*.

was tested in LB broth during daily subculture over 14 days (approximately 140 generations) (26). Strains JJ1886 and JJ1887 are very closely related strains from two sisters, one of whom had recurrent cystitis (strain JJ1887), the other of whom developed fatal urosepsis (strain JJ1886) after caring for her sister with cystitis (13). The strains share four plasmids (pJJ1886-1, pJJ1886-2, pJJ1886-3, and pJJ1886-5). After 140 generations, JJ1886 retained its five plasmids in 100% of the population, while 33% of the JJ1887 population, on average, had lost plasmid pJJ1887-4 (the equivalent of pJJ1886-5 in strain JJ1886). Although this was a short experiment in the context of experimental evolution, it demonstrates possible differences in plasmid stability even between closely related ST131-*H30Rx* strains. This may explain why some ST131-*H30Rx* strains have lost their F2:A1 plasmids. That is, during microevolution within the clade, mutational events in the chromosome and/or plasmid may create differences in plasmid stability between even closely related strains.

**The fitness cost of plasmid carriage is diminished in *H30R1* strains compared with ancestral *H30* and laboratory strains.** Some have postulated that the fitness cost for carriage of ST131 plasmids may be reduced due to epistatic interactions between plasmids and specific host strain backgrounds (27). To test this, strain competitions were performed between common reference strain K-12 MG1655 and several ST131 wild-type strains and their plasmid-containing derivatives (see Fig. S3 in the supplemental material). Using a mean competitive index as a metric, we observed two key findings. First, competitive indices in LB broth demonstrated decreasing strain fitness by sublineage, ranging from (most fit) *H30S* > *H30Rx* > *H30R1* (least fit). When the plasmids of strain JJ1886 (*H30Rx*) were introduced into different host backgrounds, competitive indices demonstrated a decreasing cost of plasmid carriage from (greatest cost) K-12 MG1655 > *H30S* > *H30R1* (least cost). In fact, no significant cost of plasmid carriage was observed in the *H30R1* genetic background. This indicates that even though overall fitness is reduced for *H30R1* and *H30Rx* compared to *H30S*, adaptive and epistatic changes may have occurred during the transition from basal clades to the *H30R1/Rx* genetic background, resulting in overall lower fitness costs for carriage of these plasmids and thus favoring plasmid retention. Further work is necessary to determine which specific traits or mutations enable these possible epistatic interactions.

**What events led to the current F plasmid distributions in ST131?** Given the evidence provided here, we propose a sequence of events leading to the current circulating plasmids within the ST131-*H30Rx* sublineage (Fig. 7). A first key event was the introduction of an F2:A1 plasmid type into either the *H22* or *H30S* sublineage basal to *H30R1* and *H30Rx*. This plasmid may or may not have contained island 1, island 2, or aerobactin-Sit. It likely did not yet contain *bla*<sub>CTX-M-15</sub>, as highly similar versions of this plasmid that lacks this gene are present in basal isolates. Ancestral ST131 clade members likely contained different F plasmid types with overlapping traits (such as



islands 1 and 2 and the Colla-containing region), including the F29:B10 plasmid type found in basal *H22* to *H35* isolates. These ancestral plasmids likely were lost before the introduction of the F2:A1 plasmid into a precursor to *H30R*. Other possibilities certainly exist. It is possible islands 1 and 2 and/or aerobactin-Sit was introduced into the F2:A1 plasmid after its introduction into the ST131 clade from other plasmid types. It is also possible that plasmid or gene transfer between clades (from recent to basal or vice versa) has confounded the inferences presented here. However, the proposed events summarized in Fig. 7 are the most parsimonious, based on our analyses.

Following introduction of the F2:A1 plasmid containing islands 1 and 2 into *H22/H30S*, our data suggest that the B1 allele of the FIB replicon was subsequently lost. The *H30R* clade then formed, serving as the hypothetical ancestor to *H30R1* and *H30Rx*. Based on our data, it is likely that an antibiotic resistance cassette containing *bla*<sub>CTX-M-15</sub> and possibly also *catB4*, *bla*<sub>OXA-1</sub>, *aac(6')Ib-cr*, and *tetAR*, was introduced via one or more IS26-mediated events. Along with accompanying chromosomal changes, this formed the *H30Rx* clade, which subsequently has been shaped extensively by IS-mediated acquisitions, deletions, and rearrangements on the plasmid. In some *H30Rx* strains, this resistance cassette was partially lost. In others, *bla*<sub>CTX-M-15</sub> integrated into the bacterial chromosome. Also, some strains have lost the plasmid entirely or key portions of it.

If we assume, based on the data, that *H30R1* and *H30Rx* have arisen in parallel from a common *H30R* ancestor and that the F2:A1 plasmid was present prior to (and, presumably, also in) this *H30R* hypothetical ancestor, this requires that an intermediate ancestor along the pathway from *H30R* to *H30R1* must have lost the F2:A1 plasmid, subsequently acquiring the F1:A2:B20 plasmid found in current *H30R1* strains. Of course, because of the challenges of interpreting sequence data from highly plastic elements, it is difficult to say to what extent recombination between plasmids impacted current plasmid formations. Given that all of the F replicon alleles were found, in some form, in *H41* and early *H22/H35* isolates, it is certainly possible that some of the current plasmids in *H30R1* and *H30Rx* arose through extensive recombination of different F-type replicons. Our data are limited in this context, as we do not have completed plasmid sequences of all possible allelic combinations.

In summary, although considerable uncertainty remains regarding precisely how the ST131 clade evolved to contain currently circulating plasmids and resistance-associated genes, this study provides some key insights. First, ST131 contains several distinct plasmid types that were acquired independently. These plasmids' evolutionary history has been challenging to interpret because they contain overlapping genetic traits, including typical F plasmid core genes and accessory islands such as islands 1 to 2 and the Colla-containing region. However, phylogenetic inference supports the independent acquisition of these traits on multiple occasions, which fits with the concept of convergent evolution toward maintaining beneficial traits on highly plastic elements. It appears both that the *H30R1/C1* and *H30Rx/C2* clades may have coadapted with these plasmids to carry them at lower cost and that the plasmids themselves are evolving toward fixation within these clades through mutational events. Given the prominence of these plasmids among ST131 sublineages, it is likely that they play multiple roles in the success of their hosts.

## MATERIALS AND METHODS

**Bacterial strains.** A previous collection of 104 diverse ST131 strains described by Price et al. was used here (5). These included strains belonging to the following *fimH* allele-based subgroups: *H22* ( $n = 26$ ), *H30S* ( $n = 5$ ), *H30R1* ( $n = 32$ ), *H30Rx* ( $n = 27$ ), *H35* ( $n = 7$ ), and *H41* ( $n = 4$ ). Draft assemblies of all isolates from this collection were screened for plasmid-associated replicons and genetic loci, as described below. From this collection, 10 strains, including at least one representative from each of the major ST131 *fimH* allele-based subgroups included in the collection, were selected for additional PacBio sequencing (Table 1). Several additional (non-ST131) reference plasmids were also used for comparative analyses (Table 1).

**DNA sequencing.** Nine strains were sequenced using PacBio technology at the Rochester Mayo Medical Genome Facility (Rochester, MN). SMRTbell template libraries were generated from previously isolated unsharred raw genomic DNA using the Pacific Biosciences SMRTbell template prep kit 1.0 (Pacific Biosciences, Menlo Park, CA). Finished DNA libraries were subsequently subjected to DNA size selection using the BluePippin DNA size selection system (Sage Science, Inc.), with a 7-kb cutoff to select

DNA fragments greater than 7 kb. Sequencing was performed on the PacBio RSII (Pacific Biosciences) using P6 polymerase binding and C4 sequencing kits, with magnetic bead loading and 240-min acquisition.

**Analysis and annotation.** Genome assemblies from PacBio data were created using HGAP 3 as part of SMRTAnalysis version 2.2. Assemblies were subjected to three rounds of polishing with Quiver. Completed and circular chromosomes or plasmids were reoriented to their replicon (for plasmids) or the origin of replication (for chromosomes) and subjected to a final round of polishing with Quiver. Genome annotation was performed using the National Center for Biotechnology Information (NCBI) Prokaryotic Genomes Automatic Annotation Pipeline (PGAAP; [http://www.ncbi.nlm.nih.gov/genome/annotation\\_prok/](http://www.ncbi.nlm.nih.gov/genome/annotation_prok/)) followed by manual curation for F-type plasmids. XPlasMap was used to construct plasmid maps (<http://www.iayork.com/Widgets.shtml#XPlasMap>). Resistance genes were identified using the CARD antibiotic resistance database (28). Plasmid types were identified using PlasmidFinder (29). F plasmid sequence types were identified using pMLST (29).

**Plasmid genome comparisons.** Comparisons of the completed sequences of ST131 plasmids with the existing collection of 104 ST131 draft genome assemblies were performed using the CGView comparison tool (30). Similarities between plasmid genomes were calculated using nucleotide BLAST with a 1,000-bp sliding window and an E value of  $1 \times 10^{-6}$ . Circular comparison maps were then generated. Completed plasmids and reference sequences were aligned using MAUVE (31). Core single nucleotide variants from this alignment were extracted from nonrepetitive regions and analyzed using maximum parsimony methods with 1,000 bootstrap replicates in MEGA5 (32).

**Plasmid profiling.** Isolation and visualization of plasmids from the 10 isolates sequenced via PacBio were performed using pulsed-field gel electrophoresis (PFGE) with S1 nuclease (33), as previously described (12), following the procedures outlined by the Centers for Disease Control and Prevention PulseNet protocols (<http://www.cdc.gov/pulsenet/pdf/ecoli-shigella-salmonella-pfge-protocol-508c.pdf>). Strain JJ1886 was used as a positive control and size standard as it contains four plasmids ranging in size from 5 to 110 kb (13).

**Plasmid transfer experiments.** Conjugal transfer of plasmids from eight donor strains into five recipient strains representing different genetic backgrounds was performed using liquid mating experiments (see Table S2 in the supplemental material). Donor and recipient strains were grown overnight in 2 ml LB broth without antibiotic selection. The next day, the donor strain was inoculated 1:100 in fresh LB broth and allowed to grow for 3 to 4 h. Donor and recipient cells were mixed at volumes of 0.2 and 1.8 ml, respectively, and incubated at 37°C without shaking for 18 h. Eighteen hours was chosen for incubation after standard pilot experiments at 3 and 6 h failed to yield transconjugants from ST131 donors (data not shown). Following incubation, cultures were vortexed heavily, serially diluted, and plated in 100- $\mu$ l aliquots onto MacConkey agar containing ampicillin (100  $\mu$ g/ml, for donor plasmid selection) with and without rifampin (100  $\mu$ g/ml, for recipient background selection). Conjugation frequencies were expressed as transconjugants per donor cell. Three replicates were performed for each strain combination.

**In vitro competitions.** Using *E. coli* K-12 strain MG1655 as a common reference, competitions were performed in LB broth against representative strains of H30S (G199), H30R1 (MVA0038), and H30Rx (JJ1886). Also included were transconjugant derivatives of the H30S and H30R1 strains containing plasmid pJJ1886-5, a prototype H30Rx plasmid from strain JJ1886. These transconjugant strains were generated by overnight matings between *E. coli* DH10B containing pJJ1886-5 as a donor strain and respective recipient strains, followed by selection on MacConkey agar containing nalidixic acid (30  $\mu$ g/ml) and ampicillin (100  $\mu$ g/ml) to identify lactose-positive transconjugant colonies. Plasmid transfer was verified using S1 nuclease PFGE, as previously described (33).

Competitions were subsequently performed by first growing strains individually overnight in 2 ml LB broth with appropriate antibiotic selection and then washing the overnight bacterial pellet twice in sterile phosphate-buffered saline. Twenty-five microliters of each competing strain was combined into 5 ml fresh LB broth. Ten-fold serial dilutions of this mixture were plated on appropriate selective media to obtain time zero counts for each strain. The mixture was then grown overnight with shaking at 37°C. The next day, 5  $\mu$ l of the overnight growth was reinoculated into 5-ml fresh LB broth and again grown overnight. Serial dilutions and plating on selective media were performed from this second overnight growth to obtain final (48-h) counts of each strain. Competitive indices for each strain competed with *E. coli* K-12 strain MG1655 were calculated as the proportion of test to reference strain counts at time zero divided by the ratio of test to reference strain counts at 2 days (34).

**Plasmid stability experiments.** The stability of plasmids from two closely related H30Rx strains, JJ1886 and JJ1887, was tested via daily passage in LB broth for 14 days following a previously described protocol (26). Briefly, overnight cultures of strains were inoculated into fresh LB broth and grown overnight at 37°C with shaking at 200 rpm. Each day, 5  $\mu$ l of overnight culture was transferred into 5 ml fresh LB broth. At days 0, 7, and 14, samples were taken from the growth, serially diluted, and plated onto LB agar without antibiotics. A PCR assay targeting genes of the plasmids of JJ1886 and JJ1887 was developed (see Table S3 in the supplemental material). PCR was performed on 90 colonies from each replicate and time point to determine the prevalence of plasmids within the culture. An ST131-specific gene was also included in the assay to rule out contamination (35). Two replicates per strain were tested.

**Accession number(s).** Completed or draft genomes have been deposited in NCBI GenBank (Table 1). PacBio raw data are available in the NCBI SRA database under BioProject no. [PRJNA307507](https://www.ncbi.nlm.nih.gov/bioproject/PRJNA307507).



## SUPPLEMENTAL MATERIAL

Supplemental material for this article may be found at <http://dx.doi.org/10.1128/mSphere.00121-16>.

- Figure S1, TIF file, 0.2 MB.
- Figure S2, TIF file, 0.2 MB.
- Figure S3, TIF file, 0.1 MB.
- Table S1, DOCX file, 0.2 MB.
- Table S2, DOCX file, 0.1 MB.
- Table S3, DOCX file, 0.1 MB.

## ACKNOWLEDGMENTS

This work was supported in part by University of Minnesota College of Veterinary Medicine funding. Computing resources were provided by the University of Minnesota Supercomputing Institute. This material also is based on work supported in part by the Office of Research and Development, Medical Research Service, Department of Veterans Affairs, grant no. 1 I01 CX000192 01 (J.R.J.), NIH grant no. R01AI106007 (E.S.), and NIH grant no. 2R21-AI117654 (L.B.P. and J.R.J.). N.S. is currently funded through a Public Health England/University of Oxford Clinical Lectureship.

The opinions expressed here are strictly those of the authors and do not necessarily reflect those of their respective institutions or the Department of Veterans Affairs.

## FUNDING INFORMATION

This work, including the efforts of James R. Johnson, was funded by Office of Research and Development, VA (1 I01 CX000192 01). This work, including the efforts of Evgeni Sokurenko and Lance B. Price, was funded by HHS | National Institutes of Health (NIH) (R01AI106007 and 2R21-AI117654).

## REFERENCES

1. Johnson JR, Tchesnokova V, Johnston B, Clabots C, Roberts PL, Billig M, Riddell K, Rogers P, Qin X, Butler-Wu S, Price LB, Aziz M, Nicolas-Chanoine MH, Debroy C, Robicsek A, Hansen G, Urban C, Platell J, Trott DJ, Zhanel G, Weissman SJ, Cookson BT, Fang FC, Limaye AP, Scholes D, Chattopadhyay S, Hooper DC, Sokurenko EV. 2013. Abrupt emergence of a single dominant multidrug-resistant strain of *Escherichia coli*. *J Infect Dis* **207**:919–928. <http://dx.doi.org/10.1093/infdis/jis933>.
2. Nicolas-Chanoine MH, Bertrand X, Madec JY. 2014. *Escherichia coli* ST131, an intriguing clonal group. *Clin Microbiol Rev* **27**:543–574. <http://dx.doi.org/10.1128/CMR.00125-13>.
3. Mathers AJ, Peirano G, Pitout JD. 2015. The role of epidemic resistance plasmids and international high-risk clones in the spread of multidrug-resistant Enterobacteriaceae. *Clin Microbiol Rev* **28**:565–591. <http://dx.doi.org/10.1128/CMR.00116-14>.
4. Schembri MA, Zakour NL, Phan MD, Forde BM, Stanton-Cook M, Beatson SA. 2015. Molecular characterization of the multidrug resistant *Escherichia coli* ST131 clone. *Pathogens* **4**:422–430. <http://dx.doi.org/10.3390/pathogens4030422>.
5. Price LB, Johnson JR, Aziz M, Clabots C, Johnston B, Tchesnokova V, Nordstrom L, Billig M, Chattopadhyay S, Stegger M, Andersen PS, Pearson T, Riddell K, Rogers P, Scholes D, Kahl B, Keim P, Sokurenko EV. 2013. The epidemic of extended-spectrum-beta-lactamase-producing *Escherichia coli* ST131 is driven by a single highly pathogenic subclone, H30-Rx. *mBio* **4**:e00377-13. <http://dx.doi.org/10.1128/mBio.00377-13>.
6. Stoesser N, Sheppard AE, Pankhurst L, De Maio N, Moore CE, Sebra R, Turner P, Anson LW, Kasarskis A, Batty EM, Kos V, Wilson DJ, Phetsouvanh R, Wyllie D, Sokurenko E, Manges AR, Johnson TJ, Price LB, Peto TE, Johnson JR, Didelot X, Walker AS, Crook DW. 2016. Evolutionary history of the global emergence of the *Escherichia coli* epidemic clone ST131. *mBio* **7**:e02162-15. <http://dx.doi.org/10.1128/mBio.02162-15>.
7. Ben Zakour NL, Alsheikh-Hussain AS, Ashcroft MM, Khanh Nhu NT, Roberts LW, Stanton-Cook M, Schembri MA, Beatson SA. 2016. Sequential acquisition of virulence and fluoroquinolone resistance has shaped the evolution of *Escherichia coli* ST131. *mBio* **7**:e00347-16. <http://dx.doi.org/10.1128/mBio.00347-16>.
8. Phan MD, Forde BM, Peters KM, Sarkar S, Hancock S, Stanton-Cook M, Ben Zakour NL, Upton M, Beatson SA, Schembri MA. 2015. Molecular characterization of a multidrug resistance IncF plasmid from the globally disseminated *Escherichia coli* ST131 clone. *PLoS One* **10**:e0122369. <http://dx.doi.org/10.1371/journal.pone.0122369>.
9. Li JJ, Spychala CN, Hu F, Sheng JF, Doi Y. 2015. Complete nucleotide sequences of *bla*(CTX-M)-harboring IncF plasmids from community-associated *Escherichia coli* strains in the United States. *Antimicrob Agents Chemother* **59**:3002–3007. <http://dx.doi.org/10.1128/AAC.04772-14>.
10. Lanza VF, de Toro M, Garcillán-Barcia MP, Mora A, Blanco J, Coque TM, de la Cruz F. 2014. Plasmid flux in *Escherichia coli* ST131 sublineages, analyzed by plasmid constellation network (PLACNET), a new method for plasmid reconstruction from whole genome sequences. *PLoS Genet* **10**:e1004766. <http://dx.doi.org/10.1371/journal.pgen.1004766>.
11. Shin J, Ko KS. 2015. Effect of plasmids harbouring *bla*CTX-M on the virulence and fitness of *Escherichia coli* ST131 isolates. *Int J Antimicrob Agents* **46**:214–218. <http://dx.doi.org/10.1016/j.ijantimicag.2015.04.012>.
12. Johnson TJ, Hargreaves M, Shaw K, Snippes P, Lynfield R, Aziz M, Price LB. 2015. Complete genome sequence of a carbapenem-resistant extraintestinal pathogenic *Escherichia coli* strain belonging to the sequence type 131 H30R subclone. *Genome Announc* **3**:e00272-15. <http://dx.doi.org/10.1128/genomeA.00272-15>.
13. Andersen PS, Stegger M, Aziz M, Contente-Cuomo T, Gibbons HS, Keim P, Sokurenko EV, Johnson JR, Price LB. 2013. Complete genome sequence of the epidemic and highly virulent CTX-M-15-producing H30-Rx subclone of *Escherichia coli* ST131. *Genome Announc* **1**:e00988-13. <http://dx.doi.org/10.1128/genomeA.00988-13>.
14. Smet A, Van Nieuwerburgh F, Vandekerckhove TT, Martel A, De-force D, Butaye P, Haesebrouck F. 2010. Complete nucleotide sequence of CTX-M-15-plasmids from clinical *Escherichia coli* isolates: insertional events of transposons and insertion sequences. *PLoS One* **5**:e11202. <http://dx.doi.org/10.1371/journal.pone.0011202>.
15. Woodford N, Carattoli A, Karisik E, Underwood A, Ellington MJ,

- Livermore DM.** 2009. Complete nucleotide sequences of plasmids pEK204, pEK499, and pEK516, encoding CTX-M enzymes in three major *Escherichia coli* lineages from the United Kingdom, all belonging to the international O25:H4-ST131 clone. *Antimicrob Agents Chemother* **53**: 4472–4482. <http://dx.doi.org/10.1128/AAC.00688-09>.
16. **Forde BM, Ben Zakour NL, Stanton-Cook M, Phan MD, Totsika M, Peters KM, Chan KG, Schembri MA, Upton M, Beatson SA.** 2014. The complete genome sequence of *Escherichia coli* EC958: a high quality reference sequence for the globally disseminated multidrug resistant *E. coli* O25b:H4-ST131 clone. *PLoS One* **9**:e104400. <http://dx.doi.org/10.1371/journal.pone.0104400>.
  17. **Chen SL, Hung CS, Xu J, Reigstad CS, Magrini V, Sabo A, Blasiar D, Bieri T, Meyer RR, Ozersky P, Armstrong JR, Fulton RS, Latreille JP, Spieth J, Hooton TM, Mardis ER, Hultgren SJ, Gordon JI.** 2006. Identification of genes subject to positive selection in uropathogenic strains of *Escherichia coli*: a comparative genomics approach. *Proc Natl Acad Sci U S A* **103**:5977–5982. <http://dx.doi.org/10.1073/pnas.0600938103>.
  18. **Wibberg D, Szczepanowski R, Eikmeyer F, Pühler A, Schlüter A.** 2013. The IncF plasmid pRSB225 isolated from a municipal wastewater treatment plant's on-site prefloder combining antibiotic resistance and putative virulence functions is highly related to virulence plasmids identified in pathogenic *E. coli* isolates. *Plasmid* **69**:127–137. <http://dx.doi.org/10.1016/j.plasmid.2012.11.001>.
  19. **Lescat M, Calteau A, Hoede C, Barbe V, Touchon M, Rocha E, Tenaille O, Médigue C, Johnson JR, Denamur E.** 2009. A module located at a chromosomal integration hot spot is responsible for the multidrug resistance of a reference strain from *Escherichia coli* clonal group A. *Antimicrob Agents Chemother* **53**:2283–2288. <http://dx.doi.org/10.1128/AAC.00123-09>.
  20. **Johnson TJ, Aziz M, Liu CM, Sokurenko E, Kisiela DI, Paul S, Andersen P, Johnson JR, Price LB.** 2016. Complete genome sequence of a CTX-M-15-producing *Escherichia coli* strain from the H30Rx subclone of sequence type 131 from a patient with recurrent urinary tract infections, closely related to a lethal urosepsis isolate from the patient's sister. *Genome Announc* **4**:e00334-16. <http://dx.doi.org/10.1128/genomeA.00334-16>.
  21. **Cusumano CK, Hung CS, Chen SL, Hultgren SJ.** 2010. Virulence plasmid harbored by uropathogenic *Escherichia coli* functions in acute stages of pathogenesis. *Infect Immun* **78**:1457–1467. <http://dx.doi.org/10.1128/IAI.01260-09>.
  22. **Johnson TJ, Siek KE, Johnson SJ, Nolan LK.** 2006. DNA sequence of a ColV plasmid and prevalence of selected plasmid-encoded virulence genes among avian *Escherichia coli* strains. *J Bacteriol* **188**:745–758. <http://dx.doi.org/10.1128/JB.188.2.745-758.2006>.
  23. **Dozois CM, Daigle F, Curtiss R III.** 2003. Identification of pathogen-specific and conserved genes expressed *in vivo* by an avian pathogenic *Escherichia coli* strain. *Proc Natl Acad Sci U S A* **100**:247–252. <http://dx.doi.org/10.1073/pnas.232686799>.
  24. **He S, Hickman AB, Varani AM, Siguier P, Chandler M, Dekker JP, Dyda F.** 2015. Insertion sequence IS26 reorganizes plasmids in clinically isolated multidrug-resistant bacteria by replicative transposition. *mBio* **6**:e00762. <http://dx.doi.org/10.1128/mBio.00762-15>.
  25. **Johnson TJ, Siek KE, Johnson SJ, Nolan LK.** 2005. DNA sequence and comparative genomics of pAPEC-O2-R, an avian pathogenic *Escherichia coli* transmissible R plasmid. *Antimicrob Agents Chemother* **49**: 4681–4688. <http://dx.doi.org/10.1128/AAC.49.11.4681-4688.2005>.
  26. **De Gelder L, Ponciano JM, Joyce P, Top EM.** 2007. Stability of a promiscuous plasmid in different hosts: no guarantee for a long-term relationship. *Microbiology* **153**:452–463. <http://dx.doi.org/10.1099/mic.0.2006/001784-0>.
  27. **Schaufler K, Semmler T, Pickard DJ, de Toro M, de la Cruz F, Wieler LH, Ewers C, Guenther S.** 2016. Carriage of extended-spectrum beta-lactamase-plasmids does not reduce fitness but enhances virulence in some strains of pandemic *E. coli* lineages. *Front Microbiol* **7**:336. <http://dx.doi.org/10.3389/fmicb.2016.00336>.
  28. **McArthur AG, Waglechner N, Nizam F, Yan A, Azad MA, Baylay AJ, Bhullar K, Canova MJ, De Pascale G, Ejim L, Kalan L, King AM, Koteva K, Morar M, Mulvey MR, O'Brien JS, Pawlowski AC, Piddock LJ, Spanogiannopoulos P, Sutherland AD, Tang I, Taylor PL, Thaker M, Wang W, Yan M, Yu T, Wright GD.** 2013. The comprehensive antibiotic resistance database. *Antimicrob Agents Chemother* **57**: 3348–3357. <http://dx.doi.org/10.1128/AAC.00419-13>.
  29. **Carattoli A, Zankari E, García-Fernández A, Voldby Larsen M, Lund O, Villa L, Møller Aarestrup F, Hasman H.** 2014. In silico detection and typing of plasmids using PlasmidFinder and plasmid multilocus sequence typing. *Antimicrob Agents Chemother* **58**:3895–3903. <http://dx.doi.org/10.1128/AAC.02412-14>.
  30. **Grant JR, Arantes AS, Stothard P.** 2012. Comparing thousands of circular genomes using the CGView comparison tool. *BMC Genomics* **13**:202. <http://dx.doi.org/10.1186/1471-2164-13-202>.
  31. **Darling AC, Mau B, Blattner FR, Perna NT.** 2004. Mauve: multiple alignment of conserved genomic sequence with rearrangements. *Genome Res* **14**:1394–1403. <http://dx.doi.org/10.1101/gr.2289704>.
  32. **Tamura K, Peterson D, Peterson N, Stecher G, Nei M, Kumar S.** 2011. MEGA5: molecular evolutionary genetics analysis using maximum likelihood, evolutionary distance, and maximum parsimony methods. *Mol Biol Evol* **28**:2731–2739. <http://dx.doi.org/10.1093/molbev/msr121>.
  33. **Barton BM, Harding GP, Zuccarelli AJ.** 1995. A general method for detecting and sizing large plasmids. *Anal Biochem* **226**:235–240. <http://dx.doi.org/10.1006/abio.1995.1220>.
  34. **Lang KS, Danzeisen JL, Xu W, Johnson TJ.** 2012. Transcriptome mapping of pAR060302, a bla<sub>CMY-2</sub>-positive broad-host-range IncA/C plasmid. *Appl Environ Microbiol* **78**:3379–3386. <http://dx.doi.org/10.1128/AEM.07199-11>.
  35. **Johnson JR, Menard M, Johnston B, Kuskowski MA, Nichol K, Zhanel GG.** 2009. Epidemic clonal groups of *Escherichia coli* as a cause of antimicrobial-resistant urinary tract infections in Canada, 2002 to 2004. *Antimicrob Agents Chemother* **53**:2733–2739. <http://dx.doi.org/10.1128/AAC.00297-09>.